MITOCHONDRIAL ANTIGEN

ATM02-02 Mitochondrial antigen 0.20 mg ATM02-10 Mitochondrial antigen 1.0 mg

Description of the Product

Purified from bovine heart muscle. After coating onto ELISA plates the product will bind autoantibodies to mitochondrial antigen.

Purity: The mitochondrial antigen (5 subunits: PDH E2 (74 kDa), PDH E3 (Protein X, 56 kDa), PDH E3 (lipoamide dehydrogenase, 55 kDa), PDH E1 α (41 kDa) and PDH E1 β (36 kDa)) is more than 90% pure, as assessed by SDS polyacrylamide gel electrophoresis.

Storage: Store at -65°C or below (long term). Avoid repeated freezing and thawing. Mix thoroughly before use.

Clinical and Biochemical Data

Antimitochondrial antibodies (AMA) were first detected in sera from patients with primary biliary cirrhosis (PBC) using immunofluorescent staining on cryostat sections of a range of tissues¹. PBC is characterised by a T-cell mediated inflammation of the medium-sized bile ducts and typically affects women over the age of forty². The mitochondrial antigens of PBC have been identified as components of a functionally related enzyme family (the 2-oxo-acid dehydrogenase complexes) of which the pyruvate dehydrogenase (PDH) complex is the most prominent autoantigenic component⁵⁻⁶. Although autoantibodies to PDH are most frequently detected in the sera of patients with PBC, they have also been reported to occur in a minority of patients with rheumatic autoimmune diseases⁷ (Sjögren's syndrome, scleroderma, SLE and rheumatoid arthritis) as well as in patients with leprosy⁸, monoclonal gammopathies⁶, tuberculosis¹⁰ or antigiomerular basement membrane disease¹¹.

2-Oxo-acid dehydrogenase complexes catalyse key reactions in intermediary metabolism^{12,13}. The PDH complex oxidatively decarboxylates pyruvate to acetyl CoA; patient autoantibodies have been reported to inhibit enzyme function^{14,15}. Each enzyme complex consists of multiple copies of three enzymes E1, E2 and E3. E2 forms a central symmetrical core to which multiple copies of E1, E3 and protein X are attached¹⁶ yielding a total complex molecular weight of several thousand kDa. As is evident from primary sequence data of the human¹⁶, pig¹⁷ and rat^{3,16} 70 kDa M2 antigen (PDH E2), that the PDH autoantigen is well conserved between species. PBC patient sera have been shown to react with PDH complexes from E. coli to human¹⁹⁻²².

The PDH complex from bovine heart muscle constitutes AroTec's mitochondrial antigen. Four of the five subunits present (E2 acetyltransferase, E3 Protein X, E1α decarboxylase and E1β decarboxylase) are known to react with AMA²³. The use of native PDH complex for the detection of AMA by ELISA has been described by several authors^{11,24-26}.

Methodology

The following is an ELISA procedure which can be used to detect antimitochondrial autoantibodies in human serum using the ATM02 purified mitochondrial antigen:

1. Dilute the purified antigen to 0.2-0.5 μ g/ml in coating buffer (20 mM Tris-HCl, pH 8.0; 0.40 M NaCl; 0.05% deoxycholate).

2. Coat ELISA plates with 100 μl of diluted antigen per well. Cover and incubate overnight at +4 °C.

3. Empty the plates and remove excess liquid by tapping on a paper towel.

4. Block excess protein binding sites by adding 200 μI PBS (10 mM potassium phosphate, pH 7.4, 0.15 M NaCl) containing 1% BSA per well. Incubate at room temperature for three hours.

5. Empty plates and apply 100 μl of serum samples diluted 1:100 in PBS / 1% BSA / 1% casein / 0.1% Tween $^{\odot}$ 20. Incubate at room temperature for 1 hour.

6. Empty plates and add 200 μ l PBS / 0.1% Tween[®] 20 per well. Incubate 5 minutes then empty plates. Repeat this step twice.

7. Apply 100 μl anti-human IgG-enzyme conjugate (horseradish peroxidase or alkaline phosphatase) diluted in PBS / 1% BSA /

1% casein / 0.1% Tween[®] 20 per well and incubate for 1 hour. 8. Repeat step 6.

9. Add enzyme substrate and stop the reaction when appropriate.

10. Read absorbance in an ELISA spectrophotometer.

References

- 1. Walker, J.G. et al. (1965) Lancet 1:827
- 2. Leung, P.S.C. et al. (1996) J. Autoimmunity 9, 119
- 3. Gershwin, M.E. et al. (1987) J. Immunol. 138, 3525
- 4. Van de Water, J. et al. (1988) J. Exp. Med. 167, 1791
- 5. Yeaman, S.J. et al. (1988) Lancet 1:1067
- 6. Coppel R.L. et al. (1995) Immunol. Rev. 144, 17
- 7. Zurgil, N. et al. (1992) J. Clin. Immunol. **12**, 201 8. Gilburd, B. et al. (1994) J. Clin. Immunol. **14**, 14
- 8. Gilburd, B. et al. (1994) J. Clin. Immunol. 14, 1
- 9. Gilburd, B. et al. (1995) Immunol. Lett. 45, 163
- 10. Klein, R. et al. (1993) Clin. Exp. Immunol. 92, 308
- 11. Marriott, J.B. et al. (1993) Clin. Exp. Immunol. 93, 259
- 12. Bassendine, M.F. et al. (1989) Seminars in Liver Dis. 9, 124 13. Behal, R.H. et al. (1993) Ann. Rev. Nutr. 13, 497
- 14. Sundin, U. (1990) Clin. Exp. Immunol. 81, 238
- 15. Teoh, K.-L. et al. (1994) Hepatology **19**, 1029
- 16. Coppel, R.C. et al. (1988) Proc. Natl. Acad. Sci. USA 85, 7317
- 17. Koike, K. Submitted Jan-2000 to EMBL/Genbank/DDBJ databases
- 18. Matuda, S. et al. (1992) Biochim, Biophys. Acta **1131**, 114
- 19. Mitchison, H.C. et al. (1986) Hepatology 6, 1279
- 20, Yeaman, S.J. et al. (1989) Biochem, J. 257, 625
- 21. Chuang, D.T. et al. (1985) J. Biol. Chem. 260, 13779
- 22. Fussey, S.P.M. et al. (1991) Hepatology 13, 467
- 23. Leung, P.S.C. et al. (1994) Man. Biol. Markers Dis. (Kluwer) B8.1
- 24. Heseltine, L. et al. (1990) Gastroenterology 99, 1786
- 25. Matsui, M. et al. (1993) Hepatology 18, 1069
- 26. Tischler, M. (1995) Clin. Exp. Rheumatol. 13, 497

NOTE: **No patented technology** has been used by AROTEC during the preparation of this product.

