

# PARIETAL CELL ANTIGEN (H/K-ATPase)

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ATP01-02 Parietal cell antigen 0.20 mg

ATP01-10 Parietal cell antigen 1.0 mg

## Description of the Product

Purified from pig gastric mucosa. After coating onto ELISA plates the product will bind anti-parietal cell antibodies specific for the H/K-ATPase  $\alpha$ - and  $\beta$ -subunits.

**Purity:** The H/K-ATPase  $\alpha$ - and  $\beta$ -subunits (95 and 60-90 kDa) are more than 90% pure, as assessed by SDS gel electrophoresis.

**Storage:** Store at -65°C or below (long term). Avoid repeated freezing and thawing. Mix thoroughly before use.

## Clinical and Biochemical Data

Autoimmune gastritis is an organ-specific autoimmune disease characterised by type A chronic atrophic gastritis and circulating autoantibodies to gastric parietal cells and intrinsic factor<sup>1</sup>. The end stage of type A chronic atrophic gastritis is pernicious anaemia, a disease which affects predominantly caucasians of north European origin and which is considered to be the most common cause of vitamin B<sub>12</sub> deficiency in Western countries<sup>2</sup>. The prevalences of pernicious anaemia and type A chronic atrophic gastritis in Western populations are estimated to be 0.1 and 1.0 % respectively<sup>2</sup>. Antiparietal cell antibodies can be detected in the serum of 80-90% of pernicious anaemia patients<sup>3</sup>.

Recent studies<sup>4,6</sup> have demonstrated that the major molecular targets recognised by parietal cell autoantibodies are the  $\alpha$ - and  $\beta$ -subunits of the gastric proton pump (H/K-ATPase)<sup>7</sup>. The H/K-ATPase is the most abundant membrane protein of the canalicular and tubovesicular membranes of parietal cells. It catalyses the countertransport of H<sup>+</sup> and K<sup>+</sup> in an electroneutral fashion at the expense of ATP and is thereby responsible for the acidification of the stomach.

Although the H/K-ATPase  $\alpha$ -subunit cDNA codes for a protein of m.wt. 114 kDa<sup>8</sup>, it migrates on SDS-electrophoresis as a protein of m.wt. 95 kDa. On the other hand, the  $\beta$ -subunit migrates as a protein of 60-90 kDa, whereas its cDNA codes for a protein of 35 kDa<sup>9</sup>. Extensive glycosylation is presumably responsible for this phenomenon<sup>10</sup>. The amino acid sequence of the human H/K-ATPase is highly homologous to that of the porcine protein<sup>9</sup>. Of the 1,035 residues of the  $\alpha$ -subunit, 1,015 are identical with those of the pig enzyme; of the 291 amino acids of the  $\beta$ -subunit, 251 are identical between the two species<sup>9</sup>. It has been demonstrated by several authors<sup>11-14</sup> that the porcine protein can serve as an effective ELISA substrate for the detection of human antiparietal cell autoantibodies.

## Methodology

The following is an ELISA procedure which can be used to detect antiparietal cell antibodies in human serum using the ATP01 purified parietal cell antigen:

1. Dilute the purified antigen to 0.5-1.0  $\mu$ g/ml in 10 mM Tris-HCl, pH 8.0 / 0.15 M NaCl / 0.1% sodium desoxycholate.
2. Coat ELISA plates with 100  $\mu$ l of diluted antigen per well. Cover and incubate overnight at +4 °C.
3. Empty plates and remove excess liquid by tapping on a paper towel.
4. Block excess protein binding sites by adding 200  $\mu$ l PBS (phosphate buffered saline) containing 1% BSA per well. Incubate at room temperature for 3 hours.
5. Empty plates and apply 100  $\mu$ l of serum samples diluted 1:100 in PBS / 1% BSA / 1% casein / 0.02% Tween<sup>®</sup> 20. Incubate at room temperature for 1 hour.
6. Empty plates and add 200  $\mu$ l PBS / 0.02% Tween<sup>®</sup> 20 per well. Incubate 5 minutes then empty plates. Repeat this step twice.
7. Apply 100  $\mu$ l anti-human IgG-enzyme conjugate (horseradish peroxidase or alkaline phosphatase) diluted in PBS / 1% BSA / 1% casein / 0.02% Tween<sup>®</sup> 20 per well and incubate for 1 hour.
8. Repeat step 6.
9. Add enzyme substrate and stop the reaction when appropriate.
10. Read absorbance in an ELISA spectrophotometer.

## References

1. Gleeson, P.A. & Toh, B.-H. (1991) *Immunol. Today* **12**, 233
2. Strickland, R. (1990) in *Immunology and Immunopathology of the Liver and Gastrointestinal Tract* (Targan, S.R. & Shanahan, F. eds.), pp. 535-546. Igaku-Shoin
3. Vandelli et al. (1979) *N. Engl. J. Med.* **300**, 1406
4. Karlsson, F.A. et al. (1988) *J. Clin. Invest.* **81**, 475
5. Burman, P. et al. (1989) *Gastroenterol.* **96**, 1434
6. Toh, B.-H. et al. (1990) *Proc. Natl. Acad. Sci. USA* **87**, 6418
7. Rabon, E.C. & Reuben, M.A. (1990) *Annu. Rev. Physiol.* **52**, 321
8. Maeda, M. et al. (1990) *J. Biol. Chem.* **265**, 9027
9. Ma, J.-Y. et al. (1991) *Biochem. Biophys. Res. Comm.* **180**, 39
10. Okamoto, C.T. et al. (1990) *Biochim. Biophys. Acta* **1037**, 360
11. Karlsson, F.A. et al. (1987) *Clin. Exp. Immunol.* **70**, 604
12. Burman, P. et al. (1991) *J. Gastroenterol.* **26**, 207
13. Chuang, J.S. et al. (1992) *Autoimmunity* **12**, 1
14. Ma, J.-Y. et al. (1994) *Scand. J. Gastroenterol.* **29**, 961

NOTE: **No patented technology** has been used by AROTEC during the preparation of this product.

